

# **Human EGF ELISA Instructions**

Cat: EH0023

## **Content**

|                         | CAT              | Volume       |
|-------------------------|------------------|--------------|
| 1 CP (Coated Plate)     | EH0023CP         | 96 well      |
| 2 S (Standard)          | EH0023S,S1~S7,S0 | 9 vial       |
| 3 DA (Detect Antibody)  | EH0023DA         | 6 ml/bottle  |
| 4 SH (Streptavidin-HRP) | ESH01            | 12 ml/bottle |
| 6 AB (Assay Buffer 1×)  | EAB01            | 12 ml/bottle |
| 6 SD (Sample Diluent)   | ESD01            | 15 ml/bottle |
| 7 TS (TMB Substrate)    | ETS01            | 12 ml/bottle |
| 8 SS (Stop Solution)    | ESS01            | 12 ml/bottle |
| 9 WB (Wash Buffer 10×)  | EWB01            | 50 ml/bottle |
| SF (Sealer Film)        | ESF01            | 6 piecse     |

**NOTE:** After the kit is opened, the stabilization period of each content is 30 days.

## REAGENT PREPARATION

### Washing Buffer (1×) Preparation

Wash Buffer  $10\times$  was diluted to  $1\times$  by glass-distilled or deionized water. Transfer to a clean wash bottle and store at 2 to  $25^{\circ}$ C.

#### Standard Curve Preparation:

S1 to S7 and S0 is ready to use for serum and plasma.

Other sample type, prepare the standard curve with whatever buffer (SPB, Sample Prepared Buffer) is used to prepare the sample, such as cell culture supernatant, tissue grinding liquid, cell lysate, etc. Urine sample use AB (Assay Buffer) prepare standard curve.

The Human EGF Standard EH0023S 10000 pg/ml 30  $\mu$ l + 270  $\mu$ l SPB serves as the high standard (1000 pg/ml). Pipette 200  $\mu$ l of SPB into each tube. Use the high standard to produce a 1:2 dilution series. Mix each tube thoroughly before the next transfer. SPB serves as the zero standard (0 pg/ml).

100µ1 100µ1 100µ1 100µ1 100µ1 100µ1

300µl

200µl

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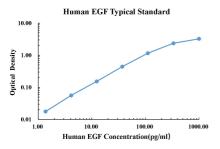
## **ASSAY PROCEDURE**

Bring all reagents and samples to room temperature before use.

- 1 Prepare all reagents and working standards as directed in the previous sections.
- 2 Remove excess CP (Coated Plate) strips from the plate frame, return them to the foil pouch and reseal.
- 3 Add 50 μl of AB (Assay Buffer) to each well.
- 4 Add 50 μl or 10 μl of Standard or sample per well. Ensure reagent addition is uninterrupted and completed within 15 minutes.
- $\bigcirc$  Add 50  $\mu$ l of  $\bigcirc$  DA (Detect Antibody) to each
- **6** Cover with an **SF** (Sealer Film). Incubate at room temperature (18 to 25°C) for 1 hours on a microplate **shaker** set at 500 rpm.
- $\begin{tabular}{ll} \begin{tabular}{ll} \beg$
- f 8 Add 100  $\mu I$  of f SH (Streptavidin-HRP) to each well
- Over with a new SF (Sealer Film). Incubate at room temperature (18 to 25°C) for 30 min on a microplate shaker set at 500 rpm.
- 10 Repeat aspiration/wash as in step 7.
- (1) Add 100  $\mu$ l of TS (TMB Substrate) to each well. Incubate for 5-30 minutes at room temperature.
- 12 Add 100 μl of SS (Stop Solution) to each well.
- (3) Determine the optical density within 30 minutes, using microplate reader set to 450 nm corrected with 570 nm or 630 nm.



## **TYPICAL DATA**



| pg/ml | O.D.  |       | Averag<br>e | Correcte<br>d |
|-------|-------|-------|-------------|---------------|
|       |       |       | c           | u             |
| 0.00  | 0.011 | 0.012 | 0.0118      |               |
| 0.00  | 5     | 1     | 0.0118      |               |
| 1.37  | 0.029 | 0.029 |             |               |
|       | 3     | 7     | 0.0295      | 0.0177        |
|       |       | /     |             | 0.01//        |
| 4.12  | 0.067 | 0.068 | 0.0680      |               |
|       | 6     | 3     | 0.0680      | 0.0562        |
| 12.35 | 0.163 | 0.167 |             |               |
|       | 3     | 5     | 0.1654      | 0.1536        |
|       |       | -     |             | 0.1336        |
| _     | 0.453 | 0.455 |             |               |

# **SENSITIVITY**

The minimum detectable dose (MDD) of human EGF is typically less than 0.12 pg/ml (50  $\mu$ l of sample volume) or 0.59 pg/ml (10  $\mu$ l of sample volume).

The MDD was determined by adding two standard deviations to the mean optical density value of ten zero standard replicates and calculating the corresponding concentration.

### **PRECISION**

- Intra-assay Precision (Precision within an assay) Three samples of known concentration were tested twenty times on one plate to assess intra-assay precision.
- Inter-assay Precision (Precision between assays)

|                                    | Intra-assay Precision |      |       | Inte | r-assay Prec | ision |       |
|------------------------------------|-----------------------|------|-------|------|--------------|-------|-------|
| Sample<br>Number                   | SI                    | S2   | 83    |      | S1           | S2    | 83    |
|                                    | 22                    | 22   | 22    |      | 6            | 6     | 6     |
| Average<br>(pg/ml)                 | 13.2                  | 61.3 | 217.8 |      | 20.4         | 105.0 | 347.9 |
| Standard<br>deviation              | 0.5                   | 4.1  | 6.6   |      | 1.0          | 5.3   | 19.3  |
| Coefficient of<br>variation<br>(%) | 3.5                   | 6.7  | 3.0   |      | 4.9          | 5.1   | 5.6   |

## **RECOVERY**

The spike recovery was evaluated by spiking 3 levels of human EGF into health human serum sample. The un-spiked serum was used as blank in this experiment.

The recovery ranged from 78% to 106% with an overall mean recovery of 93%.

#### LINEARITY

To assess the linearity of the assay, five samples were spiked with high concentration of EGF in human serum and diluted with Sample Diluent to produce samples with values within the dynamic range of the assay.

The linearity ranged from 87% to 101% with an overall mean recovery of 95%.

### SAMPLE VALUES

Serum/Plasma – Thirty samples from apparently healthy volunteers were evaluated for the presence of EGF in this assay. No medical histories were available for the donors.

| Sample<br>Matrix | Sample<br>Evaluated | Range<br>(pg/ml) | Detectable<br>% | Mean of<br>Detectable<br>(pg/ml) |
|------------------|---------------------|------------------|-----------------|----------------------------------|
| Serum            | 30                  | 200.7-417.0      | 100             | 304.2                            |

n.d. = non-detectable. Samples measured below the sensitivity are considered to be non-detectable.